

GENERAL ICC-P TROUBLESHOOTING TIPS

SYMPTOM	ISSUE	RECOMMENDATIONS
	<div>Antibody Application</div>	Increase the concentration or incubation time of the primary or secondary antibody.
	<div>Permeabilization Buffer</div>	Use the proper permeabilization reagent for the target protein's localization. Triton detergent is necessary for mitochondrial or nuclear proteins, but will dissolve the outer membrane and disrupt proper membrane localization.
	<div>Cell Fixation</div>	Increase incubation duration or detergent concentration.
	<div>Antibody Compatibility</div>	Over fixation can cause epitope masking. Decrease the time or concentration of the fixative.
	<div>Target Availability</div>	Confirm that your primary and secondary antibodies are compatible by checking the species reactivity.
	<div>Cells Drying</div>	Confirm the antibody can be used for assays in which the protein is in its native conformation.
	<div>Cell Viability</div>	Ensure that the secondary is working and compatible with your microscope's filter sets by using a positive control primary.
	<div>Microscope Adjustments</div>	Use an overexpression assay or positive control cell line known to express the protein of interest.
		Fluorescent signal will be lost if the cells are allowed to dry. Ring coverslips with nail polish.
 Background	<div>Antibody Concentration</div>	Confirm cell viability before starting the staining procedure.
	<div>Blocking</div>	Increase the exposure time of your camera.
	<div>Antibody Application</div>	Decrease the concentration of the primary/secondary antibody.
	<div>Contamination Artifacts</div>	Increase the incubation time or concentration of serum in the blocking buffer. Use blocking buffer for primary and secondary antibody dilutions.
	<div>Washing</div>	Always incubate primary antibodies overnight at 4° C. Room temperature incubation increases unspecific binding and causes higher background.
	<div>Spectral Overlap</div>	Confirm that the secondary is not crossreacting with the cells by performing the assay without the primary.
		Ensure slides are clean and free of dust. Buffers should be made fresh to prevent microbial contamination.
	<div>Washing</div>	Increase the amount of washes. Add very gentle agitation to the plates.
	<div>Spectral Overlap</div>	Increase the concentration of Tween in the PBS-T.
		If double or triple labeling the cells, confirm that the secondaries do not overlap into the same spectral range.