

## **05/19/2016 Immunoprecipitation test for pol $\beta$ antibody (Novus Biologicals, NB100 – 91734)**

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### **A. Materials used:**

1. DNA Polymerase beta antibody (NB100-91734) purchased from Novus Biologicals, USA.
2. **10X Cell Lysis Buffer** from Cell Signaling Technology (CAT#9803): To prepare 10 ml of 1X cell lysis buffer, add 1 ml 10X cell lysis buffer to 9 ml Milli Q water, mix. Add 1 mM PMSF and proteinase inhibitor cocktail immediately prior to use.
3. **Protein A Agarose Beads from Cell Signaling Technology (CAT# 9863).**
4. **Normal rabbit IgG from Santa Cruz Biotechnology (CAT# sc-2027).**
5. **Human U2OS cells (ATCC# HTB-96) cultured in DMEM + 10% FBS + pen/Strep.**
6. **Purified recombinant human DNA polymerase  $\beta$  (His-tagged) were prepared by the Demple Lab, Department of Pharmacological Sciences, Stony Brook University, Stony Brook, New York).**

### **B. Preparing Cell Lysates**

1. To harvest cells, remove media and rinse cells once with ice-cold 1X PBS.
2. Remove PBS and add 0.4 ml ice-cold 1X cell lysis buffer to each plate (10 cm) and incubate on ice for 5 min.
3. Scrape cells off the plate and transfer to microcentrifuge tubes. Gently rock the suspension on an orbital shaker at 4°C for 15 minutes to lyse cells.
4. Microcentrifuge for 15 min at 4°C, 14,000 x g and transfer the supernatant to a new tube. The supernatant is the cell lysate.

### **C. Immunoprecipitation**

1. **Protein A agarose beads were washed by PBS and restored to 50% slurry with PBS.**
2. **To pre-clear cell lysates**, 20  $\mu$ l of 50% protein A bead slurry were added to 200  $\mu$ l cell lysate at 0.6 mg/ml.
3. Incubate with rotation at 4°C for 30 min.
4. Microcentrifuge for 10 min at 4°C. Transfer the supernatant to a fresh tube.
5. DNA polymerase $\beta$  antibody (Novus biologicals, NB100-91734) was added to 200  $\mu$ l of pre-cleared cell lysate at 0.6 mg/ml to final antibody concentration of 2 $\mu$ g/ml.
7. Incubate for 2 hours on a rotating wheel at 4°C.
8. Add 20 $\mu$ l of protein A beads (50% bead slurry), incubate with gentle rotating for 2 hr at 4°C.
9. Microcentrifuge for 30 sec at 4°C. Save supernatants for control loading and wash pellet five times with 500  $\mu$ l of 1X cell lysis buffer. Keep on ice between washes.

#### D. Sample Analysis by Western Immunoblotting

1. Resuspended the pellet in 20  $\mu$ l of 2X SDS sample buffer.
2. Samples were boiled for 5 min and microcentrifuge for 1 min at 14,000 x g.
3. Load samples (including supernatant and input samples) onto a 12% gel for SDS-PAGE.
4. Analyze sample by western blot.

#### E. Result:

It turned out that this antibody is not suitable for IP application, as shown on the image:

- 1) NB100-91734 antibody didn't pulldown endogenous DNA polymerase  $\beta$  from cell lysates (lane 7), which is the same as the normal rabbit IgG control (lane 6). It is believed that DNA polymerase  $\beta$  protein stayed in the supernatant fraction, although lane 2 and lane 3 didn't show the polymerase  $\beta$  band, that is due to the fact that 10% of supernatant just equal to 12ug of cell lysate, which is below the detectable level in view of the fact that only 30ug of cell lysate could show a specific endogenous polymerase  $\beta$  band (lane 10).
- 2) NB100-91734 antibody did pulled down a portion of purified recombinant human polymerase  $\beta$  (His-tagged) (lane 8 and 9), but its binding capacity is very limited, seeing that 10% supernatant already showed pretty strong bands in both 100ng and 25ng IP reactions (lane 4 and 5).

#### Conclusion:

CAT# NB100-91734, DNA polymerase  $\beta$  antibody is not suitable for immunoprecipitation use.

05/19/2016 DNA pol $\beta$  antibody (NB100-91734) IP testing

